

SUPPLEMENTARY INFORMATION

Supplementary Methods

Plasmids and shRNA sequences

A TEV protease cleavage site (ENLYFQG) followed by a FLAG tag was introduced via PCR into the APP695 sequence and subcloned in peak12-APP (Lichtenthaler et al., 2003) with PmlI/NotI to obtain peak12-APP-TEV-FLAG. peak12 vectors expressing APPs β , APPs β ' or APPs α ending in amino acids 14, 15 or 16 of the A β sequence were amplified via PCR and subcloned into PmlI/NotI digested peak12-APP vector. HindIII/NotI digested APP-TEV-FLAG was subcloned into HindIII/NotI digested pCR8/GW/TOPO-pCS2-MCS to obtain pCR8/GW/TOPO-APP-TEV-FLAG. FU- Δ Zeo was generated by digestion of FU-Valentin vector (Lois et al., 2002) with BstZ17I/PMEI followed by religation. FU- Δ Zeo-DsRed-UAS-GW was generated by subcloning the SnaBI/NheI digested DsRed-UAS-GW cassette of pT2D-Dest_pA_DsRed.T4_E1b_UAS_E1b_GW-R1-R2_pA provided by Christian Haass (Paquet et al., 2009) into AfeI/NheI digested FU- Δ Zeo vector. FU- Δ Zeo-Gal4-VP16 was generated by PCR amplification of Gal4-VP16 from pT2D-Exp_EF1a_Gal4VP16_pA provided by Christian Haass (Paquet et al., 2009). The PCR product was subcloned into NheI/NotI digested FU- Δ Zeo vector. FU- Δ Zeo-DsRed-UAS-APP-TEV-FLAG was generated by Gateway cloning L/R recombination between the donor plasmid pCR8-GW-TOPO-APP-TEV-FLAG and FU- Δ Zeo-DsRed-UAS-GW. pLVTHM was described (Wiznerowicz and Trono, 2003). To obtain pLVTHMmod the H1 element was amplified by PCR and subcloned into pLVTHM vector with BamHI/XmaI. shRNA oligo duplexes targeting ADAM10 (AD10) were ligated into MluI/XmaI digested pLVTHMmod vector to obtain lentiviral knockdown plasmids pLVTHMmod-Control, -sh6, -sh7, -sh9, -MM8. Sh9 sequence was adapted from the most potent siRNA of the Dharmacon pool. pLKO2mod-EGFP-WPRE was generated by replacing the hairpin of pLKO1-mmADAM10-TRC44 (RNAi consortium) with an MluI/XmaI site and a subsequent replacement of the puromycin resistance gene with EGFP-WPRE. shRNA oligo duplexes targeting murine (mmA10) and human AD10, murine AD9, murine AD17 and human BACE1 (B1) were ligated into MluI/XmaI digested pLKO2mod-EGFP-WPRE vector to obtain pLKO2mod-EGFP-WPRE-mmA10-sh1, mmA10-sh2, B1-sh1, B1-Sh2, mmA17-sh1, mmA9-sh1. pcDNA 3.1(-)-VSV-G was generated by digestion of pHCMV-VSV-G with SalI/EagI which was subcloned into XhoI/EagI digested pcDNA3.1(-). psPAX2 was from Didier Trono (Addgene Plasmid 12260).

Supplementary References

- Lichtenthaler, S.F., Dominguez, D.I., Westmeyer, G.G., Reiss, K., Haass, C., Saftig, P., De Strooper, B. and Seed, B. (2003) The cell adhesion protein P-selectin glycoprotein ligand-1 is a substrate for the aspartyl protease BACE1. *J Biol Chem*, **278**, 48713-48719.
- Lois, C., Hong, E.J., Pease, S., Brown, E.J. and Baltimore, D. (2002) Germline transmission and tissue-specific expression of transgenes delivered by lentiviral vectors. *Science*, **295**, 868-872.
- Paquet, D., Bhat, R., Sydow, A., Mandelkow, E.M., Berg, S., Hellberg, S., Falting, J., Distel, M., Koster, R.W., Schmid, B. and Haass, C. (2009) A zebrafish model of tauopathy allows in vivo imaging of neuronal cell death and drug evaluation. *J Clin Invest*, **119**, 1382-1395.
- Wiznerowicz, M. and Trono, D. (2003) Conditional suppression of cellular genes: lentivirus vector-mediated drug-inducible RNA interference. *J Virol*, **77**, 8957-8961.

Supplementary Table 1: shRNA sequences

Name:	shRNA sequence
Con	5'-CCCCaacaagatgaagagcaccaaTTCAAGAGAttggtgctcttcattctgttg TTTTGGAAA-3'
sh6	5'-CCCCaagttgcctcctcctaaccactTTCAAGAGAGtggttaggaggaggcaactTTTTTGGAAA-3'
sh7	5'-CCCCgacatttcaacctacgaattTTCAAGAGAAaattcgtagggtgaaatgctTTTTTGGAAA-3'
sh9	5'-CCCCggacaaacttaacaacaatTTCAAGAGAAattgttgaagttgtccTTTTTGGAAA-3'
B1-sh1	5'-CCCCggtacaaagactgcgtcttgattcaagagatcaagacgcagctcttgtaccTTTTTGGAAA-3'
B1-sh2	5'-CCCCgcgtgacagaaacagagaaattctcaagagagattctctgtctgtcacgcTTTTTGGAAA-3'
A10-sh1	5'-CCGGtagacattatgaaggattatcctcgaggataatccttcataatgtctaTTTTTGGAAA-3'
A10-sh2	5'-CCGGcagacttggtctcgcataaacctcgaggtttatcgagagccaagtctgTTTTTGGAAA-3'
shA9	5'-CCGGgctcctgcacctcctttatatctcgagatataaaggaggtgcaggagcTTTTTGGAAA-3'
shA17	5'CCCCgcagcactccataaggaaactcgagttccttatggagtgcctgcTTTTTGGAAA-3'

Sequences sh6, sh7 and sh9 are against human ADAM10, A10-sh1 and A10-sh2 against murine ADAM10. shA9 is against murine ADAM9, shA17 against murine ADAM17 and B1-sh1 and B1-sh2 are against human BACE1. Sh7 has been described previously (Reiss et al., 2005).

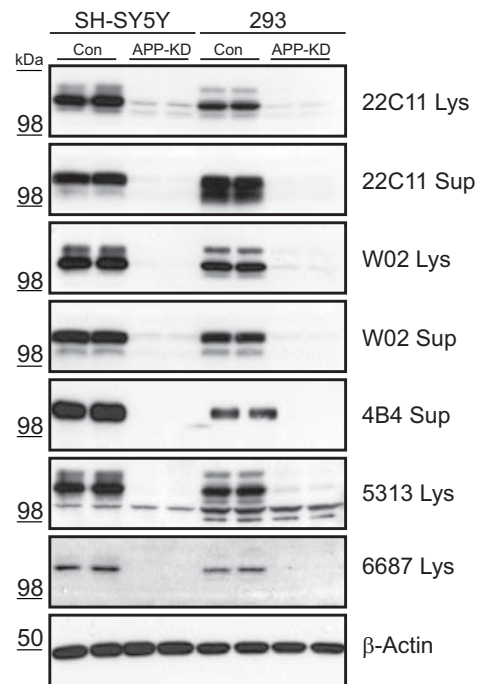
Supplementary Table 2: Primer sequences for quantitative RT-PCR

Genes	Primer sequences	Product size (bp)
ADAM9	For 5'-GTTTACACCTACGACAAGGAAGGC-3' Rev 5'-TCCCTCCACATAGCCTCGATAGT-3'	87
ADAM10	For 5'-TAAGGAATTATGCCATGTTTGCTGC-3' Rev 5'-ACTGAACTGCTTGCTCCACTGCA-3'	91
ADAM17	For 5'-TTGGAGCAGAACATGACCCTGATGG-3' Rev 5'-TGCAGCAGGTGTCGTTGTTTCAGGTA-3'	276
Actb	For 5'-ATGCTCCCCGGGCTGTAT-3' Rev 5'-CATAGGAGTCCTTCTGACCCATTC-3'	87
GapDH	For 5'-TGTGTCCGTCGTGGATCTGA-3' Rev 5'-TTGCTGTTGAAGTCGCAGGAG-3'	150
Tbp	For 5'-GGCCTCTCAGAAGCATCACTA-3' Rev 5'-GCCAAGCCCTGAGCATAA-3'	107

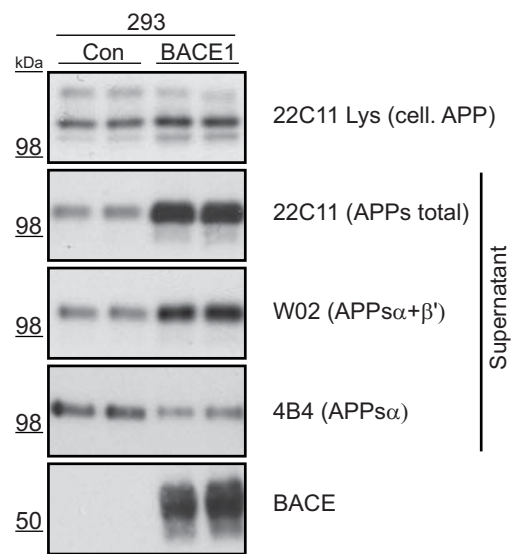
Suppl. Fig. S1: Specificity of APP antibodies. HEK293 and SH-SY5Y cells expressing endogenous APP were transfected with a control siRNA pool (Con) or a siRNA pool against APP (APP-KD). Cell lysates (Lys) were analyzed for APP with 22C11, 6687, 5313 and W02. Conditioned media (Sup) were analyzed for APP with 22C11, W02 and 4B4. As a loading control, cell lysates were probed for β -actin. APP was efficiently knocked-down. The blots show the specificity of the antibodies for APP.

Suppl. Fig. S2: Overexpression of the β -secretase BACE1 reduces 4B4 signal. Conditioned media and cell lysates of HEK293 cells either transfected with an empty control vector or BACE1 were analyzed for endogenous cellular APP (22C11), α + β '-cleaved APP (W02), α -cleaved APP (4B4) and BACE1 as transfection control. BACE1 transfection reduced α -secretase cleavage and 4B4 signal, as expected.

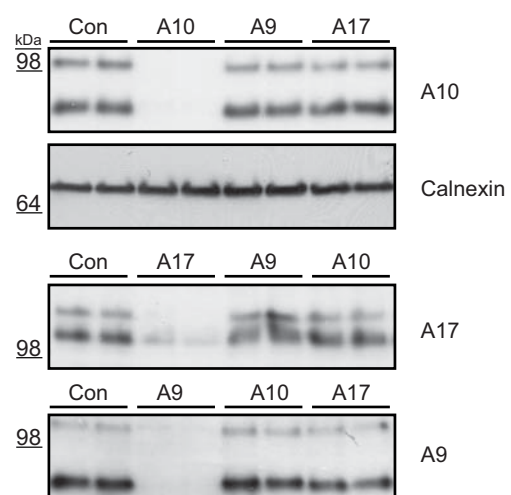
Suppl. Fig. S3: Knockdown of either ADAM9, 10 or 17 does not alter the protein levels of immature and mature forms of the other proteases. Membrane preparations of SH-SY5Y cells either treated with siRNA pools against ADAM9, 10, 17 (A9, A10, A17) or a control siRNA pool (Con) were probed for ADAM9, 10 and 17. As loading control membranes were analyzed for calnexin.



Suppl. Fig.S1



Suppl. Fig. S2



Suppl. Fig.S3